

Bioremediation of Heavy Metals (Pb and Hg) from Contaminated Sites by Potential Micro-organisms

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Received: 20 Dec 2023; Revised accepted: 27 Feb 2024; Published online: 13 Mar 2024

Abstract

Contemporary apprehensions about environmental degradation stem from various natural and anthropogenic activities, encompassing rapid industrialization, global economic expansion, increased agrochemical use, and increased hydrocarbon deposition. This multifaceted array of activities contributes to environmental decay, resulting in decreased crop yields and deleterious effects on the biota. Heavy metals emerge as prominent contaminants in environmental pollution, characterized by their nonbiodegradable nature and prolonged persistence in ecosystems. The deleterious repercussions of heavy metal contamination on soil and aquatic organisms are multiple, thereby necessitating vigilant consideration. The adverse consequences of heavy metal exposure extend beyond the environmental impact to include significant implications for human health. Consequently, remedial interventions become imperative to eliminate hazardous organic heavy metals or facilitate their conversion into less harmful inorganic forms. This review synthesizes the existing literature on the intricate interplay between anthropogenic activities, heavy metal contamination, and the resulting environmental and health implications. By scrutinizing these intricate dynamics, this review seeks to illuminate the imperatives of effective remediation strategies in mitigating the adverse consequences of heavy metal pollution, thus fostering sustainable environmental practices and protecting public health.

Key words: Bioremediation, Contaminants, Mercury, Lead, Microorganisms

Significant aspects of life are air, soil, and water, and they are polluted by various natural and anthropogenic activities [1]. Natural activities such as volcanic eruption, erosion, and rock disintegration are the main sources of pollution [2]. Anthropogenic activities, such as active urbanization, increasing global economy and industrialization [1]. Human activities cause degradation in soil quality due to heavy metal contamination; various heavy metals present in the soil such as mercury (Hg), cadmium (Cd), lead (Pb), arsenic (As) and chlorium (Cr) cause toxicity to living organisms [3]. Elements with the highest density of more than 4-5 g/cm³ present on earth are known as heavy metals and even in low concentrations they pose toxicity, classified as extremely poisonous inorganic chemicals [4]. The heavy metals present in the environment pose a serious threat to living organisms and the ecosystem [5-9]. Heavy metals possess the ability of bioaccumulation and non-biodegradability, which is of great concern to the environment [10-11]. Containment due to heavy metals in the environment poses a serious health hazard such as heart disease, liver damage, neurological diseases, cardiovascular diseases, cancer, hypophosphatemia, and damage to the central nervous system [12]. Heavy metals can accumulate in various cell organelles such as mitochondria, lysosomes, endoplasmic reticulum and the process of protein formation, which has a toxic effect on the biological system [13].

Industrially lead is one of the trace elements that are referred to as toxicant, occupational hazard, environmental pollutant, and biological nutrient [14]. There are various sources of lead discharge in the environment, such as plating, mining, lead smelting, storage batteries, the ceramic and glass industries, and the manufacturing of tetraethyl lead [15]. Since the prehistoric ages, lead has been used in various fields, such as casting, piping, ammunition, and building materials [16]. Along with the use of lead, it can also cause damage to wildlife, as lead is one of the major causes of the destruction of wildlife and causes damage to human health such as anaemia, damage to the nervous system, and kidney diseases [17]. Higher concentrations of lead exposure during pregnancy can lead to various serious problems such as miscarriage and sterility in women, while a high concentration of lead in men affects increased blood lead levels of up to 1.9 µmol/l which destroy semen quality. Therefore, lead is classified as a carcinogen that causes cancer [18]. To prevent such health hazards, eco-friendly and cost-effective techniques should be used to remove toxic heavy metals and their effects.

Mercury is a very harmful metal, and methyl mercury causes a wide range of health hazards such as neurological damage, paralysis, blindness, chromosome damage, irritability, insanity, and birth defects [19-20]. Mostly environmental mercury is present in two forms such as organomercuric salt

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Citation: Kulkarni SS, Narsinge AP, Chavan RS, Mali GV. 2024. Bioremediation of heavy metals (Pb and Hg) from contaminated sites by potential micro-organisms. *Res. Jr. Agril. Sci.* 15(2): 368-375.

and inorganic mercury, the most common species of mercuric salt are HgS, HgCl₂, CH₃HgCl and CH₃HgOH [21]. Most organomercuric compounds are derived from methyl mercury, which is one of the very toxic species because it accumulates quickly in tissues of biological systems [22]. Mercury is released from various sources, natural and anthropogenic activities, and some are reemission-originating [23-24]. Various anthropogenic activities of mercury emission are the mining, various agricultural materials, combustion, and industrial discharge, which annually releases about 2000 to 2200 tons of mercury [25-27].

Various physicochemical methods have been used to remove and treat environmental contaminants, such as heavy metals. Methods include evaporation, sorption, precipitation, ion exchange, reverse osmosis, and electrochemical treatment [28-29]. However, these physicochemical methods have their disadvantages such as high reagent, partial removal of metals, higher use of energy, production of toxic waste or by-products [30]. Therefore, to overcome such problems, cost-effective and environmentally friendly alternative tools are necessary [31]. Utilizing naturally occurring materials such as clays, zeolites, activated carbon, and biochar as sorbents or adsorbents can effectively remove contaminants from water and soil. These

materials are abundant, cost-effective, and often exhibit high selectivity and efficiency in pollutant removal.

Bioremediation is one of the processes that uses various microbes such as bacteria, fungi, algae, and higher plants to treat heavy metal pollution [32]. Bioremediation tools are used to convert toxic heavy metals into their less toxic form [33-35]. The presence of excessive heavy metals in the environment causes microorganisms to evolve several mechanisms such as adsorption, chemical reduction, and complexation to survive in a polluted environment and to clean up the polluted environment [36-37].

Industrial sources and effects of heavy metals (Hg and Pb)

Metals are naturally present in the environment, but some natural as well as anthropogenic activities lead to excess emission of heavy metals, worldwide lead emission in the atmosphere is 12.0 thousand tons per year naturally, while 332.0 thousand tons per year through anthropogenic activities [38]. The main sources of lead emission are coal burning, mining, automobile emissions, smoking, paint and sources of mercury emissions are batteries, the paper industry, and pesticides [31]. The effect of heavy metals on human health shown in (Table 1).

Table 1 Effect of heavy metal mercury and lead on human health and plants

Heavy metal	Effects on human health	Effects on plant	References
Mercury	Loss of memory, kidney damage, lung cancer, autoimmune disease, insomnia, fatigue, depression.	Inhibit mitochondrial activity and generate oxidative stress leading to the formation of reactive oxygen species (ROS), interfere with cellular metabolism	[39-40]
Lead	Cardiovascular diseases, short-term memory loss, problems in learning and coordination.	In plants, it reduces germination, decreases cell division, and impairs mineral nutrition.	[41-43]

Microbial bioremediation of Hg and Pb

Microorganisms can alter the drastically changing environment and sustain themselves in the presence of inorganic heavy metals. To adapt to changing environment microorganisms, use various mechanisms such as the production of exopolysaccharide (EPS), exclusion, enzyme use, biotransformation and metallothionein production to survive the toxicity of metals [44-45]. The mechanism evolved by microorganisms for resistance to various heavy metals and detoxification of heavy metals includes redox processes, ion exchange, precipitation, and surface complexation [46]. The most common mechanisms used by microbes to resist heavy metals are the production of biosurfactant, metallothionein, metal oxidation, metal-ligand degradation, methylation, demethylation, metal efflux pump, extracellular and intracellular sequestration of metals, exclusion of the permeability barrier, and enzymatic decrease [47]. Microorganisms carry a negative charge on the surface of the cell, and this anionic charge helps the microorganism bind to the metal cation, which facilitates the adsorption mechanism [48].

The absorption of heavy metals by various microorganisms occurs via two mechanisms such as bioaccumulation, which is an active process, whereas adsorption is a passive transport. Various microorganisms, such as bacteria, algae, plants, and fungi, are used to decontaminate or clean up the heavy metal-polluted environment [49-50].

Various microorganisms have different biosorption abilities that differ among a wide range of microbe biomass, and these microorganisms adjust to altered physical and chemical environments to increase biosorption activity [51]. Bacteria are one of the important biosorbents with the ability to survive in extreme conditions and to resist heavy [52-53].

Marine bacteria are extensively resistant to the concentration of 25 ppm mercury, which is one of the highly toxic heavy metals, marine mercury-resistant bacteria are capable of cleaning up not only mercury but also cadmium and lead. These marine bacteria identified using 16s rRNA are *Bacillus pumilus*, *Brevibacterium iodinium*, *Pseudomonas aeruginosa*, and *Bacillus spp.* Within 96 hours. These bacteria remove about 98% of Pb which shows a good detoxification efficiency and can be used in the implementation of these bacteria for the bioremediation of heavy metals [54]. The study shows *Vibrio fluvialis* has the capacity to remove mercury with an efficient reduction of 60% mercury ions at a concentration of 250 µg/ml. The strain was isolated from the discharge of effluent from the SIPCOT industrial area and identified using 16s r RNA. *V. fluvialis* shows great bioremediation activity and minimal antibiotic resistance. Therefore, mercury was successfully removed in an environmentally friendly way [55].

Various microorganisms such as bacteria, yeast and protozoa isolated from the tannery effluent were *Bacillus licheniformis*, *Candida parapsilosis*, and *Tetrahymena rostrata*. They showed mercury-resistant activity was identified using 16 s r RNA and 18 s r RNA. These strains successfully demonstrated mercury removal individually and in consortia. When these strains were processed individually, *Bacillus licheniformis*, *Candida parapsilosis*, and *Tetrahymena rostrata* showed a capacity of mercury removal of 73%, 80%, and 40%, respectively, when strains used in combination such as *Bacillus licheniformis* and *Candida parapsilosis* minimize the concentration to 85%, the combination of *C. parapsilosis* and *T. rostrata* reduced 77% mercury, while 73% mercury removed when *B. licheniformis* and *T. rostrata* were used. The consortia of these 3 microorganisms successfully removed 88% mercury after 96 h of incubation [56]. The two locally isolated strains

Klebsiella pneumoniae and *Pseudomonas aeruginosa* showed mercury biosorption activity with significant pH. *K. pneumoniae* showed the optimal biosorption activity at pH 5 with 15% efficiency, while *Pseudomonas aeruginosa* showed the highest biosorption activity at pH 5.8 with almost 25% efficiency [57]. The tannery industries are the main source of heavy metal emissions such as lead (Pb), chromium (Cr), and cadmium (Cd), three strains were screened *Gamella sp.*, *Micrococcus sp.* and *Hafnia sp.* shown degradation potential in which *Gamella* effectively showed a reduction in Pb of $55.16 \pm 0.06\%$ and $36.55 \pm 0.01\%$ shown by *Micrococcus sp.* [1]. One of the studies deals with the removal of Pb with various

heavy metals by *Bacillus firmus*, various parameters checked during the study were the initial concentration of metal ions, the concentration of polysaccharides, the pH of the solution based on the optimal pH parameter 98.3% of the maximum amount of Pb was removed by *Bacillus* [58]. During the textile industry dye effluent study, various strains of bacteria such as *B. licheniformis*, *P. fluorescens*, *E. coli*, and *S. typhi* were isolated and studied on three heavy metals such as Cadmium, Lead and zinc in which *Pseudomonas fluorescens* removed the maximum lead 94.32%, 92.15% by *Salmonella typhi*, 89.45% by *Bacillus licheniformis* and 86.73% by *E. coli* [59]. The degradation capacity efficacy in percentage shown in (Table 2).

Table 2 Heavy metals degradation efficacy of various bacteria

Type of the Microorganisms	Name of microorganisms	Heavy metal	Biosorption efficiency	References
Bacteria	<i>Bacillus pumilus</i> , <i>Brevibacterium iodinium</i> , <i>Pseudomonas aeruginosa</i> , and <i>Bacillus spp.</i>	Pb	98%	[54]
	<i>Vibrio fluvialis</i>	Hg	60%	[55]
	<i>Bacillus licheniformis</i>	Hg	73%	[56]
	<i>Klebsiella pneumoniae</i>	Hg	15%	[57]
	<i>Pseudomonas aeruginosa</i>	Hg	25%	[57]
Consortia of organisms	<i>B. licheniformis</i> and <i>C. parapsilosis</i>	Hg	85%	[56]
	<i>C. parapsilosis</i> and <i>T. rostrata</i>		77%	
	<i>B. licheniformis</i> and <i>T. rostrata</i>		73%	
	<i>B. licheniformis</i> , <i>C. parapsilosis</i> and <i>T. rostrata</i>		88%	
Bacteria	<i>Gamella sp.</i>	Pb	$55.16 \pm 0.06\%$	[1]
	<i>Micrococcus sp.</i>	Pb	$36.55 \pm 0.01\%$	[1]
	<i>Bacillus firmus</i>	Pb	98.3%	[58]
	<i>Pseudomonas fluorescens</i>	Pb	94.32%	[59]
	<i>Salmonella typhi</i>	Pb	92.15%	[59]
Bacteria	<i>Bacillus licheniformis</i>	Pb	89.45%	[59]
Bacteria	<i>E. coli</i>	Pb	86.73%	[59]
Protozoa	<i>Tetrahymena rostrata</i>	Hg	40%	[56]

Bioremediation of mercury and lead by algae

Heavy metals can be removed using algae, which has one of the greatest abilities for the sorption of heavy metals [60]. The possible mechanism for the removal of heavy metals was absorption and storage of heavy metals, which differs from the type, anatomy of algae, to the conditions of the growing medium [62]. The best alternative today is the use of algae for bioremediation purposes, and it has a great capacity to produce biodiesel, on the surface of algae heavy metals can be easily adsorbed, in this study *Chlorella vulgaris* was used for the biosorption of lead (Pb²⁺) and its removal efficiency was 99.4% [7]. During the studies by researchers, *Anabaena sp.*, *Trichodesmium sp.*, *Oscillatoria sp.*, *Cylindrospermopsis sp.* and *Nostoc sp.* removed the maximum lead. Showed a minimum inhibitory concentration to various heavy metals, but *Nostoc sp.* removed the maximum lead. Up to 99.6%, therefore, *Nostoc* can be used in future bioremediation [61]. In another study that confirmed the potential for enhanced sorption capacity, the cross-linking of native biomass of *A. nodosum* with bis(ethenyl)sulfone involved the incorporation of sulfone groups into the biosorbent material. Cross-linked *Ascophyllum nodosum* demonstrated substantial lead uptake, reaching levels as high as 370 P mg/g [62]. The research suggests that the readily accessible and cost-effective green marine algae *C. fascicularis* can serve as an effective biosorbent material for the removal of Pb (II) from wastewater. Achieving a maximum adsorption capacity of 198.5 mg/g at 298 K and pH 5.0 demonstrates its efficiency in this regard [63]. The objective of

this study was to investigate the potential utilization of *Cladophora glomerata* biomass as an economical sorbent for the removal of heavy metal Pb (II) ions from aqueous solutions. Through batch experiments, it was evident that *Cladophora glomerata* exhibited a noteworthy capacity to adsorb heavy metal ions from Pb(II). The maximum biosorption capacity for these ions in *Cladophora glomerata* was determined to be 25.5 mg/g. The findings indicated that the biosorbent's uptake capacity was influenced by both the pH and the initial metal concentration. Furthermore, modifications involving acid/heat positively influenced biosorption yield and metal uptake [64]. *Oscillatoria quadripunctulata* demonstrated an effective capacity to remove lead from both sewage and petrochemical industry effluent, ranging from 32% to 100% [65]. The dried biomass of *Spirogyra hyalina* was used as a biosorbent for the removal of cadmium (Cd), mercury (Hg), lead (Pb), arsenic (As) and cobalt (Co) from aqueous solutions, considering various initial concentrations of the heavy metals and contact times. By utilizing *Spirogyra hyalina* biomass as a biosorbent, this approach offers a sustainable and environmentally friendly solution for heavy metal removal from aqueous solutions, potentially mitigating the adverse effects of metal contamination on ecosystems and human health. The findings revealed that the highest adsorption of Cd, Hg, and as occurred when the initial concentration of these heavy metals was established at 40 mg/l. In the case of Pb, metal uptake demonstrated an upward trend with increasing initial concentration, showing the lowest uptake at 20 mg/l (q=5.495)

within 30 minutes of contact time and the highest at 80 mg/l (q=15.471) after 120 minutes [66-67]. Various researchers use

algae to clean up the contaminated environment, as shown in the (Table 3).

Table 3 Heavy metals degradation efficacy of various algae

Algae	Heavy metal	Biosorption efficiency (%) and maximum uptake capacity (mg/g)	Reference
<i>Chlorella vulgaris</i>	Lead	99.4 %	[10]
<i>Nostoc spp.</i>	Lead	99.6 %	[61]
<i>Ascophyllum nodosum*</i>	Lead	78%, 72%	[62-63]
<i>Cladophora fascicularis</i>	Pb (II)	198.5 mg/g	[63]
<i>Cladophora glomerata**</i>	Pb	22.5 mg/g	[64]
<i>Oscillatoria quadripunctulata</i>	Pb	32-100%	[65]
<i>Sargassum natans, Sargassum vulgare</i>	Pb	214 mg/g	[62]
<i>Spirogyra hyaline</i>	Hg, Pb	Hg, (40 mg/g); Pb (80 mg/g)	[67]

*Algal species cross-linked with other chemicals at pH 4.5 and 3.5

**Biosorption capacity at 4.0 pH

Bioremediation of mercury and lead by plants

Phytoremediation is one of the promising eco-friendly and economical techniques derived from plants that has been used as an alternative for bioremediation for many years [68-69]. Phytoremediation is primarily a technique that uses plant and correlated microorganisms to remediate contaminants to clean up the polluted environment, such as wastewater, groundwater, and sludge [44]. Almost the whole plant system can accumulate essential and non-essential metals that play no significant role [70]. A wide variety of wild plants can grow in polluted areas that contain heavy metals that accumulate in the roots and shoots. Heavy metal Pb accumulates in the roots of

Brassica, which is transported to the shoot; therefore, Brassica is used to decontaminate the environment, called phytoextraction [71]. Various plants grown in industrial areas can accumulate heavy metals; this study was effectively carried out on Pb, the plants that accumulate Pb are *E. cheiradenia*, *Scariola orientalis*, *Centaurea virgata*, *Gundelia tournefortii* and *Eleagnum angustifolia* [72]. In one of the studies, the *X. pensylvanicum* plant was used for the removal of various heavy metals, but the strong activity shown at pH 4 and below after 90 min of shaking time showed that the maximum heavy metal removed was Pb [73]. The more research on bioremediation of mercury and lead by plants shown in (Table 4).

Table 4 Bioremediation of mercury and lead by plants

Plant species	Heavy metal	Reference
Brassica juncea	Pb	[71]
<i>Euphorbia cheiradenia, Scariola orientalis, Centaurea virgata, Gundelia tournefortii</i> and <i>Eleagnum angustifolia</i>	Pb	[72]
<i>Xanthium pensylvanicum</i>	Pb	[73]
<i>Thlaspi caerulescens, Alyssum lesbiacum</i>	Pb	[74]
<i>Jatropha curcas</i>	Pb	[75]
<i>Populus deltoides, Populus nigra, Populus trichocarpa</i>	Pb	[76]
<i>Populus deltoids</i>	Hg	[77]
<i>Trifolium alexandrinum</i>	Pb	[78]
<i>Zea mays</i>	Pb	[79]
<i>Zea mays, Ambrosia artemisiifolia</i>	Pb	[80]

Bioremediation of mercury and lead by fungi

With the wide variety of microorganisms implemented in the biosorption of various heavy metals, fungi have also shown bioaccumulation and biosorption activity in an eco-friendly manner, and today fungi are used as the best alternative to bacteria [81]. Fungi appear to be hyperactive accumulators, as fungi are generally accompanied in areas rich in heavy metals [82]. For the reduction of heavy metal pollution, fungi are one of the best organisms used to monitor studies that can be directly exposed to heavy metals in the environment [83]. The individual and combined effectiveness of microorganisms in the processing of mercury was assessed. When used separately, *Candida parapsilosis* demonstrated the ability to remove 80% of mercury [56]. The research investigated the biosorption of Hg (2+) by *Saccharomyces cerevisiae* (SC). The experimental findings indicate that SC serves as an effective biosorbent for Hg~(2+). Under conditions where pH is set at 3 and temperature is 15 ° C, using 40g / L biomass of SC as a biosorbent for a 0.5g/L Hg (2+) solution of 0.5g / L of Hg (2+) results in a remarkable 96% removal rate of Hg (2+). The adsorption process is rapid and equilibrium is reached in 15 minutes [84]. A bioremediation method utilizing *Aspergillus niger* was

devised to generate mild organic acids, facilitating the leaching of heavy metals from polluted soils. The fungus was cultured on the surfaces of three distinct contaminated soils (clay loam, loam, and sandy clay loam) for 15 days at a temperature of 30 ° C and a pH below 4. In the case of clay loam, the leaching process resulted in a reduction of 85% in mercury levels [85]. In this study, we isolated *Aspergillus terreus*, a fungal strain with demonstrated tolerance, from the enriched effluent. Subsequent individual testing of these isolates revealed significant biosorption capabilities for metal ions. In particular, the fungal isolate exhibited a remarkable 97.13% biosorption of Pb after 144 hours, underscoring its promising potential for bioremediation [86]. Similarly to the investigation, previous research has documented the biosorption capacity for Pb2+ of the live biomass of *Mucor rouxii* treated with NaOH, revealing a capacity of 36.69 mg/g [87]. The soil in the troughs is regulated to a pH of 5.5. The study reveals varying percentages of Pd2+ removal, ranging from 67% to 82%. Specifically, the removal efficiency reached 82% for 5 ppm, 76% for 10 ppm, 73% for 25 ppm, 67% for 50 ppm and 67% for 100 ppm concentrations [88]. Bioremediation of Pb by *Saprolegnia delica* was influenced by temperature, with optimal

performance observed at 20°C, resulting in the elimination of 90% of the initial Pb concentration. As the temperature increased, the bio removal efficiency of *Saprolegnia delica* for Pb followed this order: 25°C > 30°C > 35°C (with removal percentages of 82.5%, 68.8% and 24.6% of the total Pb

concentration, respectively). The least bio removal of Pb by *Saprolegnia delica* occurred at 15°C, where only 18.8% of the initial Pb was sequestered [89]. Heavy metals degradation efficacy of various fungi with their percent degradation efficacy was shown in (Table 5).

Table 5 Heavy metals degradation efficacy of various fungi

Name of the microorganisms	Bio remediator	Heavy metal	Biosorption efficiency (%) and maximum uptake capacity (mg/g)	References
Fungi	<i>Candida parapsilosis</i>	Hg	80%	[56]
Fungi	<i>Saccharomyces cerevisiae</i>	Hg	55.76 (mg/g)	[84]
Fungi	<i>Aspergillus niger</i>	Pb	85%	[85]
Fungi	<i>Aspergillus terreus</i>	Pb	97.13%	[86]
Fungi	<i>Mucor rouxii</i>	Pb	36.69 mg/g	[87]
Fungi	<i>Saccharomyces cerevisiae</i>	Pb	67-82%	[88]
Fungi	<i>Trichoderma viridae</i> , <i>Saprolegnia delica</i>	Pb (II)	100% and 90%	[89]

CONCLUSION

This review article mainly focuses on the bioremediation of heavy metals lead and mercury. The current state of heavy metal bioremediation, examined in this study, offers great potential for metal detoxification. The cell wall of the biosorbents contains peptidoglycans and polysaccharides that act as active binding sites for increased metal uptake. Faster kinetics, high metal binding over a wide pH range, and

temperature are just a few benefits of this technique, which is also environmentally friendly and cost-effective. This review offers a chance to discuss how microbial cells and their metabolites contribute to heavy metal remediation and environmental research. These would make it easier to create more effective methods for heavy metal bioremediation in the environment.

Conflict of interest: None

Funding: None

LITERATURE CITED

- Marzan LW, Hossain M, Mina SA, Akter Y, Chowdhury AMA. 2017. Isolation and biochemical characterization of heavy-metal resistant bacteria from tannery effluent in Chittagong city, Bangladesh: Bioremediation viewpoint. *The Egyptian Journal of Aquatic Research* 43(1): 65-74.
- Yadav R, Kumar R, Gupta RK, Kaur T, Yodha K, Kour A, Kaur S, Rajput A. 2023. Heavy metal toxicity in earthworms and its environmental implications: A review. *Environmental Advances*. pp 100374.
- Su C. 2014. A review on heavy metal contamination in the soil worldwide: Situation, impact and remediation techniques. *Environmental Skeptics and Critics* 3(2): 24.
- Hoyle-Gardner J, Badisa V, Ibeanusi V, Mwashote B, Jones W, Brown A. 2020. Application of innovative bioremediation technique using bacteria for sustainable environmental restoration of soils from heavy metals pollution: A review. *Journal of Bioremediation and Biodegradation* 11(3): 1-6.
- Deepa C, Suresha S. 2014. Biosorption of lead (II) from aqueous solution and industrial effluent by using leaves of *Araucaria cookii*: Application of response surface methodology. *IOSR Journal of Environmental Science, Toxicology and Food Technology* 8(7): 67-79.
- Hryniewicz K, Baum C. 2014. Application of microorganisms in bioremediation of environment from heavy metals. *Environmental Deterioration and Human Health: Natural and Anthropogenic Determinants*. pp 215-227.
- Igiri BE, Okoduwa SI, Idoko GO, Akabuogu EP, Adeyi AO, Ejiogu IK. 2018. Toxicity and bioremediation of heavy metals contaminated ecosystem from tannery wastewater: A review. *Journal of Toxicology* 2018: 2568038.
- Okolo N, Olowolafe E, Akawu I, Okoduwa S. 2016. Effects of industrial effluents on soil resources in Challawa industrial area, Kano, Nigeria. *Journal of Global Ecology and Environment* 5(1): 1-10.
- Siddiquee S, Rovina K, Azad SA, Naher L, Suryani S, Chaikaew P. 2015. Heavy metal contaminants removal from wastewater using the potential filamentous fungi biomass: A review. *Jr. Microb. Biochem. Technology* 7(6): 384-395.
- Aung WL, Hlaing NN, Aye N. 2013. Biosorption of lead (Pb 2+) by using *Chlorella vulgaris*. *International Journal of Chemical, Environmental and Biological Sciences* 1(2): 2320-4087.
- Gautam RK, Soni S, Chattopadhyaya MC. 2015. Functionalized magnetic nanoparticles for environmental remediation. *In: Handbook of research on diverse applications of nanotechnology in biomedicine, chemistry, and engineering*. pp 518-551. IGI Global.
- Pandey AK, Shrivastava A. 2018. Bioremediation of lead contaminated soil using bacteria. *Res. Jr. Life Sci. Bioinform. Chem. Science* 4: 355-361.
- Wang S, Shi X. 2001. Molecular mechanisms of metal toxicity and carcinogenesis. *Molecular and Cellular Biochemistry* 222: 3-9.
- Khan H, Jamaluddin Ahmed M, Iqbal Bhangar M. 2006. A simple spectrophotometric method for the determination of trace level lead in biological samples in the presence of aqueous micellar solutions. *Spectroscopy* 20(5/6): 285-297.
- Rajendra Sukhadeorao D. 2020. Lead: Toxicological profile, pollution aspects and remedial solutions. *In: (Eds) C. Pipat. Lead Chemistry* (pp. Ch. 3). IntechOpen. <https://doi.org/10.5772/intechopen.93095>
- Congeevaram S, Dhanarani S, Park J, Dexilin M, Thamaraiselvi K. 2007. Biosorption of chromium and nickel by heavy metal resistant fungal and bacterial isolates. *Journal of Hazardous Materials* 146(1/2): 270-277.

17. Iranzo M, Sainz-Padro I, Boluda R, Sanchez J, Mormeneo S. 2001. The use of microorganisms in environmental engineering. *Annals of Microbiology* 51: 135-143.
18. Lindbohm ML, Sallmén M. 2017. Reproductive effects caused by chemical and biological agents. Finnish Institute of Occupational Health.
19. Fergusson JE. 1990. *The Heavy Elements: Chemistry, Environmental Impact and Health Effects*. Pergamon Press, Oxford. pp 85-547.
20. Luckey TD, Venugopal B. 1978. *Metal Toxicity in Mammals: Chemical Toxicity of Metals and Metalloids*. B. Venugopal and TD Luckey. Plenum Press.
21. Al-Sulaiti MM, Soubra L, Al-Ghouti MA. 2022. The causes and effects of mercury and methylmercury contamination in the marine environment: A review. *Current Pollution Reports* 8(3): 249-272.
22. Munthe J, Kindbom K, Parsmo R, Yaramenka K. 2019. *Technical Background Report to the Global Mercury Assessment 2018*. In: IVL Svenska Miljöinstitutet.
23. Panagos P, Jiskra M, Borrelli P, Liakos L, Ballabio C. 2021. Mercury in European topsoils: Anthropogenic sources, stocks and fluxes. *Environmental Research* 201: 111556.
24. Sonke JE, Angot H, Zhang Y, Poulain A, Björn E, Schartup A. 2023. Global change effects on biogeochemical mercury cycling. *Ambio* 52(5): 853-876.
25. Ballabio C, Jiskra M, Osterwalder S, Borrelli P, Montanarella L, Panagos P. 2021. A spatial assessment of mercury content in the European Union topsoil. *Science of the Total Environment* 769: 144755.
26. Singh S, Dhyani S, Pujari PR. 2022. Coal-fired thermal power plants and mercury risks: Status and impacts to realize minamata convention promises. *Anthropocene Science* 1(4): 419-427.
27. Zheng X, Cao H, Liu B, Zhang M, Zhang C, Chen P, Yang B. 2022. Effects of mercury contamination on microbial diversity of different kinds of soil. *Microorganisms* 10(5): 977.
28. Guidi Nissim W, Palm E, Mancuso S, Azzarello E. 2018. Trace element phytoextraction from contaminated soil: A case study under Mediterranean climate. *Environmental Science and Pollution Research* 25: 9114-9131.
29. Xiong J, Wu L, Tu S, Van Nostrand JD, He Z, Zhou J, Wang G. 2010. Microbial communities and functional genes associated with soil arsenic contamination and the rhizosphere of the arsenic-hyperaccumulating plant *Pteris vittata* L. *Applied and Environmental Microbiology* 76(21): 7277-7284.
30. Ahalya N, Ramachandra T, Kanamadi R. 2003. Biosorption of heavy metals. *Research Journal of Chemistry and Environment* 7(4): 71-79.
31. Alluri HK, Ronda SR, Settalluri VS, Bondili JS, Suryanarayana V, Venkateshwar P. 2007. Biosorption: An eco-friendly alternative for heavy metal removal. *African Journal of Biotechnology* 6(25).
32. Gupta A, Joia J, Sood A, Sood R, Sidhu C, Kaur G. 2016. Microbes as potential tool for remediation of heavy metals: a review. *Jr. Microb. Biochem. Technology* 8(4): 364-372.
33. Abbas SH, Ismail IM, Mostafa TM, Sulaymon AH. 2014. Biosorption of heavy metals: A review. *Jr. Chem. Science and Technology* 3(4): 74-102.
34. Akcil A, Erust C, Ozdemiroglu S, Fonti V, Beolchini F. 2015. A review of approaches and techniques used in aquatic contaminated sediments: metal removal and stabilization by chemical and biotechnological processes. *Journal of Cleaner Production* 86: 24-36.
35. Ndeddy Aka RJ, Babalola OO. 2016. Effect of bacterial inoculation of strains of *Pseudomonas aeruginosa*, *Alcaligenes fecalis* and *Bacillus subtilis* on germination, growth and heavy metal (Cd, Cr, and Ni) uptake of *Brassica juncea*. *International Journal of Phytoremediation* 18(2): 200-209.
36. Mohammed AS, Kapri A, Goel R. 2011. Heavy metal pollution: source, impact, and remedies. *Biomanagement of Metal-Contaminated Soils*. pp 1-28.
37. Okoduwa SIR, Igiri B, Udeh CB, Edenta C, Gauje B. 2017. Tannery effluent treatment by yeast species isolates from watermelon. *Toxics* 5(1): 6.
38. Clarke A, Johnston NM. 2003. Antarctic marine benthic diversity. *Oceanography And Marine Biology* 41: 47-114.
39. Gulati K, Banerjee B, Lall SB, Ray A. 2010. Effects of diesel exhaust, heavy metals and pesticides on various organ systems: Possible mechanisms and strategies for prevention and treatment. *Indian Jr. Exp. Biology* 48(7): 710-721.
40. Messer RL, Lockwood PE, Tseng WY, Edwards K, Shaw M, Caughman GB, Lewis JB, Wataha JC. 2005. Mercury (II) alters mitochondrial activity of monocytes at sublethal doses via oxidative stress mechanisms. *Journal of Biomedical Materials Research Part B: Applied Biomaterials* 75(2): 257-263.
41. Abdul G. 2010. Effect of lead toxicity on growth, chlorophyll and lead (Pb⁺) contents of two varieties of maize (*Zea mays* L.). *Pakistan Journal of Nutrition* 9(9): 887-891.
42. Padmavathiamma PK, Li LY. 2007. Phytoremediation technology: Hyper-accumulation metals in plants. *Water, Air, and Soil Pollution* 184: 105-126.
43. Wuana RA, Okieimen FE. 2011. Heavy metals in contaminated soils: A review of sources, chemistry, risks and best available strategies for remediation. *Communications in Soil Science and Plant Analysis* 42: 111-122.
44. Dixit R, Wasiullah X, Malaviya D, Pandiyan K, Singh UB, Sahu A, Shukla R, Singh BP, Rai JP, Sharma PK. 2015. Bioremediation of heavy metals from soil and aquatic environment: An overview of principles and criteria of fundamental processes. *Sustainability* 7(2): 2189-2212.
45. Wu G, Kang H, Zhang X, Shao H, Chu L, Ruan C. 2010. A critical review on the bio-removal of hazardous heavy metals from contaminated soils: issues, progress, eco-environmental concerns and opportunities. *Journal of Hazardous Materials* 174(1/3): 1-8.
46. Yang T, Chen ML, Wang JH. 2015. Genetic and chemical modification of cells for selective separation and analysis of heavy metals of biological or environmental significance. *TrAC Trends in Analytical Chemistry* 66: 90-102.

47. Ramasamy K, Kamaludeen, Banu SP. 2007. Bioremediation of metals: microbial processes and techniques. *Environmental Bioremediation Technologies* 173-187.
48. Gavrilescu M. 2004. Removal of heavy metals from the environment by biosorption. *Engineering in Life Sciences* 4(3): 219-232.
49. Kim IH, Choi JH, Joo JO, Kim YK, Choi JW, Oh BK. 2015. Development of a microbe-zeolite carrier for the effective elimination of heavy metals from seawater. *Journal of Microbiology and Biotechnology* 25(9): 1542-1546.
50. Singh N, Verma T, Gaur R. 2013. Detoxification of hexavalent chromium by an indigenous facultative anaerobic *Bacillus cereus* strain isolated from tannery effluent. *African Journal of Biotechnology* 12(10): 1091-1103.
51. Fomina M, Gadd GM. 2014. Biosorption: current perspectives on concept, definition and application. *Bioresource Technology* 160: 3-14.
52. Srivastava S, Agrawal S, Mondal M. 2015. A review on progress of heavy metal removal using adsorbents of microbial and plant origin. *Environmental Science and Pollution Research* 22: 15386-15415.
53. Wang J, Chen C. 2009. Biosorbents for heavy metals removal and their future. *Biotechnology Advances* 27(2): 195-226.
54. De J, Ramaiah N, Vardanyan L. 2008. Detoxification of toxic heavy metals by marine bacteria highly resistant to mercury. *Marine Biotechnology* 10: 471-477.
55. Saranya K, Sundaramanickam A, Shekhar S, Swaminathan S, Balasubramanian T. 2017. Bioremediation of mercury by *Vibrio fluvialis* screened from industrial effluents. *BioMed Research International* 2017: 6509648.
56. Muneer B, Iqbal M, Shakoori F, Shakoori A. 2013. Tolerance and biosorption of mercury by microbial consortia: Potential use in bioremediation of wastewater. *Pakistan Journal of Zoology* 45(1): 247-254.
57. Al-Garni SM, Ghanem KM, Ibrahim AS. 2010. Biosorption of mercury by capsulated and slime layerforming Gram-ve bacilli from an aqueous solution. *African Journal of Biotechnology* 9(38): 6413-6421.
58. Salehizadeh H, Shojaosadati S. 2003. Removal of metal ions from aqueous solution by polysaccharide produced from *Bacillus firmus*. *Water Research* 37(17): 4231-4235.
59. Basha SA, Rajaganesh K. 2014. Microbial bioremediation of heavy metals from textile industry dye effluents using isolated bacterial strains. *Int. Jr. Curr. Microbiol. Appl. Science* 3: 785-794.
60. Mehta S, Gaur J. 2005. Use of algae for removing heavy metal ions from wastewater: progress and prospects. *Critical Reviews in Biotechnology* 25(3): 113-152.
61. Kumaran NS, Sundaramanicam A, Bragadeeswaran S. 2011. Adsorption studies on heavy metals by isolated cyanobacterial strain (*Nostoc sp.*) from uppanar estuarine water, southeast coast of India. *Journal of Applied Sciences Research* 7(11): 1609-1615.
62. Holan Z, Volesky B. 1994. Biosorption of lead and nickel by biomass of marine algae. *Biotechnology and Bioengineering* 43(11): 1001-1009.
63. Deng L, Su Y, Su H, Wang X, Zhu X. 2007. Sorption and desorption of lead (II) from wastewater by green algae *Cladophora fascicularis*. *Journal of Hazardous Materials* 143(1/2): 220-225.
64. Yalçın E, Çavuşoğlu K, Maraş M, Bıyıkoğlu M. 2008. Biosorption of lead (II) and copper (II) metal ions on *Cladophora glomerata* (L.) Kütz.(Chlorophyta) algae: Effect of algal surface modification. *Open Chemistry* 16: 689-701.
65. Halder S. 2014. Bioremediation of heavy metals through fresh water microalgae: A review. *Scholars Academic Journal of Biosciences* 2(11): 825-830.
66. Kumar JN, Oommen C. 2012. Removal of heavy metals by biosorption using freshwater alga *Spirogyra hyalina*. *Journal of Environmental Biology* 33(1): 27.
67. Mane P, Bhosle A, Jangam C, Vishwakarma C. 2011. Bioadsorption of selenium by pretreated algal biomass. *Adv. Appl. Sci. Research* 2(2): 202-207.
68. Chaney RL, Malik M, Li YM, Brown SL, Brewer EP, Angle JS, Baker AJ. 1997. Phytoremediation of soil metals. *Current Opinion in Biotechnology* 8(3): 279-284.
69. Gleba D, Borisjuk NV, Borisjuk LG, Kneer R, Poulev A, Skarzhinskaya M, Dushenkov S, Logendra S, Gleba YY, Raskin I. 1999. Use of plant roots for phytoremediation and molecular farming. *Proceedings of the National Academy of Sciences* 96(11): 5973-5977.
70. Djingova R, Kuleff I. 2000. Instrumental techniques for trace analysis. *Trace Metals in the Environment* 4: 137-185. Elsevier.
71. Kumar PN, Dushenkov V, Motto H, Raskin I. 1995. Phytoextraction: the use of plants to remove heavy metals from soils. *Environmental Science and Technology* 29(5): 1232-1238.
72. Chehregani A, Malayeri BE. 2007. Removal of heavy metals by native accumulator plants. *International Journal of Agriculture and Biology* 9(3): 462-465.
73. Salehzadeh J. 2013. Removal of heavy metals Pb 2, Cu 2, Zn 2, Cd 2, Ni 2, Co2 and Fe 3 from aqueous solutions by using *Xanthium pensylvanicum*. *Leonardo Jr. Science* 23: 97.
74. Baker A, Mcgrath S, Reeves R. 1991. In situ decontamination of heavy metal polluted soils using crops of metal-accumulating plants: A feasibility study. International Symposium on In Situ and On Site Bioreclamation, San Diego, March 1991.
75. Abhilash P, Jamil S, Singh N. 2009. Transgenic plants for enhanced biodegradation and phytoremediation of organic xenobiotics. *Biotechnology Advances* 27(4): 474-488.
76. Ruttens A, Boulet J, Weyens N, Smeets K, Adriaensen K, Meers E, Van Slycken S, Tack F, Meiresonne L, Thewys T. 2011. Short rotation coppice culture of willows and poplars as energy crops on metal contaminated agricultural soils. *International Journal of Phytoremediation* 13(Sup1): 194-207.
77. Che D, Meagher RB, Heaton AC, Lima A, Rugh CL, Merkle SA. 2003. Expression of mercuric ion reductase in Eastern cottonwood (*Populus deltoides*) confers mercuric ion reduction and resistance. *Plant Biotechnology Journal* 1(4): 311-319.
78. Ali H, Naseer M, Sajad MA. 2012. Phytoremediation of heavy metals by *Trifolium alexandrinum*. *International Journal of Environmental Sciences* 2(3): 1459-1469.

79. Meers E, Van Slycken S, Adriaensen K, Ruttens A, Vangronsveld J, Du Laing G, Witters N, Thewys T, Tack F. 2010. The use of bio-energy crops (*Zea mays*) for 'phytoattenuation' of heavy metals on moderately contaminated soils: A field experiment. *Chemosphere* 78(1): 35-41.
80. Huang J, Cunningham S. 1996. Lead phytoextraction: species variation in lead uptake and translocation. *New Phytologist* 134(1): 75-84.
81. Rajapaksha R, Tobor-Kaplon M, Baath E. 2004. Metal toxicity affects fungal and bacterial activities in soil differently. *Applied and Environmental Microbiology* 70(5): 2966-2973.
82. Purvis O, Halls C. 1996. A review of lichens in metal-enriched environments. *The Lichenologist* 28(6): 571-601.
83. Garty J. 2001. Biomonitoring atmospheric heavy metals with lichens: theory and application. *Critical Reviews in Plant Sciences* 20(4): 309-371.
84. Madrid Y, Cabrera C, Perez-Corona T, Camara C. 1995. Speciation of methylmercury and Hg (II) using baker's yeast biomass (*Saccharomyces cerevisiae*). Determination by continuous flow mercury cold vapor generation atomic absorption spectrometry. *Analytical Chemistry* 67(4): 750-754.
85. Wasay SA, Barrington SF, Tokunaga S. 1998. Using *Aspergillus niger* to bioremediate soils contaminated by heavy metals. *Bioremediation Journal* 2(3/4): 183-190.
86. Zango UU, Muhammad I, Sharma V, Sharma AK. 2023. Effective bioremediation of Cd, Cr, and Pb in tannery effluent using *Aspergillus fumigatus* and *Aspergillus terreus*: Synergistic effects of using the two strains together. *Water, Air and Soil Pollution* 234(12): 735.
87. Faryal R, Sultan A, Tahir F, Ahmed S, Hameed A. 2007. Biosorption of lead by indigenous fungal strains. *Pakistan Journal of Botany* 39(2): 615.
88. Damodaran D, Suresh G, Mohan R. 2011. Bioremediation of soil by removing heavy metals using *Saccharomyces cerevisiae*. 2nd International Conference on Environmental Science and Technology, Singapore,
89. Ali EH, Hashem M. 2007. Removal efficiency of the heavy metals Zn (II), Pb (II) and Cd (II) by *Saprolegnia delica* and *Trichoderma viride* at different pH values and temperature degrees. *Mycobiology* 35(3): 135-144.